

Neurobehavioral Effects of Developmental Methylmercury Exposure

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Methylmercury (MeHg) is a global environmental problem and is listed by the International Program of Chemical Safety as one of the six most dangerous chemicals in the world's environment. Human exposure to MeHg primarily occurs through the consumption of contaminated food such as fish, although catastrophic exposures due to industrial pollution have occurred. The fetus is particularly sensitive to MeHg exposure and adverse effects on infant development have been associated with levels of exposure that result in few, if any, signs of maternal clinical illness or toxicity. High levels of prenatal exposure in humans result in neurobehavioral effects such as cerebral palsy and severe mental retardation. Prenatal exposure to MeHg in communities with chronic low-level exposure is related to decreased birthweight and early sensorimotor dysfunction such as delayed onset of walking. Neurobehavioral alterations have also been documented in studies with nonhuman primates and rodents. Available information on the developmental neurotoxic effects of MeHg, particularly the neurobehavioral effects, indicates that the fetus and infant are more sensitive to adverse effects of MeHg. It is therefore recommended that pregnant women and women of childbearing age be strongly advised to limit their exposure to potential sources of MeHg. Based on results from human and animal studies on the developmental neurotoxic effects of methylmercury, the accepted reference dose should be lowered to 0.025 to 0.06 MeHg $\mu\text{g}/\text{kg}/\text{day}$. Continued research on the neurotoxic effects associated with low level developmental exposure is needed. — *Environ Health Perspect* 103(Suppl 6):135–142 (1995)

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Introduction

It is well established that prenatal exposure to certain toxic chemicals can have profound and irreversible effects on the physical and mental development of children. Of agents known to act as toxicants, those that cause central nervous system damage (neurotoxicants) constitute a particularly significant public health hazard. While episodes of high-dose neurotoxicant exposure to children have shown clear evidence of neurological disorders, the more subtle action of moderate to low-dose neurotoxicant exposure on important parameters of behavioral development is increasingly evident (1–3). Laboratory-based research and documented incidents of human exposure to neurotoxicants such as lead, alcohol, and methylmercury (MeHg) point to a continuum of effects in exposed children ranging

from subtle behavioral changes to frank expression of neurological damage and death. In general, the long-term consequences of developmental exposure to neurotoxicants are only now beginning to receive scientific attention.

Although there are a number of important neurotoxicants, this article focuses on the effects of developmental exposure to MeHg, a prevalent environmental contaminant. The serious public health concerns associated with MeHg exposure have resulted in world-wide attention, research, and review (4–14). The aim of this article is to provide an updated overview of the developmental effects of MeHg exposure, reexamine currently recommended exposure guidelines, and highlight future research needs.

In recognition of the adverse effects of MeHg exposure, state and federal government agencies and international agencies have developed recommendations to limit MeHg exposure. For example, the U.S. Food and Drug Administration (U.S. FDA) recommends a limit of 1 $\mu\text{g}/\text{g}$ (1 ppm) mercury in the edible portion of fish. The U.S. Environmental Protection Agency (U.S. EPA) has established a reference dose (RfD) for methylmercury at 0.3 $\mu\text{g}/\text{kg}/\text{day}$, which is equivalent to consumption of 19 μg per day of MeHg for a 62-kg woman (15). The RfD is defined as an estimate of a daily exposure to a human

population that is likely to be without an appreciable risk of deleterious effects during a life time and is meant to include sensitive subgroups. The RfD typically is arrived at through standard risk assessment procedures that include a careful evaluation of study results, determination of the lowest observed adverse effect level (LOAEL) dose, and division of the LOAEL by an appropriate uncertainty factor to yield a RfD. The World Health Organization (WHO) does not use the RfD nomenclature but developed a similar recommendation that is equivalent to 0.47 $\mu\text{g}/\text{kg}/\text{day}$ in adults (16) while noting that pregnant women, nursing mothers, and their infants are likely to be at greater risk. A recent reevaluation of the RfD for MeHg was done by Stern (14), who concluded that the RfD should be lowered to 0.07 $\mu\text{g}/\text{kg}/\text{day}$ based on human and animal studies on developmental effects of MeHg. Upon review of the human and animal literature, the authors of the current review reached a similar conclusion.

Mercury Contamination — Methylmercury Exposure

Mercury is generally released into the environment in an inorganic form by both natural and anthropogenic sources (4,11,16,17). Natural sources of mercury include emissions from volcanoes, degassing of the earth's crust, and evaporation from water.

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Artificial sources of mercury include industrial pollution, burning of fossil fuels, mining, refuse incineration, and cremation. Natural emissions of mercury into the atmosphere have been estimated to range from 2700 to 6000 metric tons per year, while world wide mining of mercury is estimated to yield 10,000 tons per year (4). It is estimated that human activities result in the release of 3000 tons per year of mercury. A significant source of environmental mercury contamination is gold mining in countries such as in Brazil (18,19). Mercury release due to gold mining results in environmental contamination through direct effluent discharge into local waterways and through volatilization (18). Approximately 3 to 5 kg of mercury are used to extract 1 kg of gold, of which 25 to 45% is lost during the amalgamation process (20). Atmospheric mercury undergoes photochemical oxidation and is then scavenged by atmospheric particulates or precipitation. Global contamination occurs as the mercury is washed out of the atmosphere onto soil, vegetation, and water (4,21).

The majority of environmental MeHg contamination has occurred through the biotransformation of inorganic mercury to organic mercury (MeHg) in a process termed methylation. During methylation, inorganic mercury is converted into MeHg by microbial action, primarily in sediments of fresh and ocean waters. Methylmercury readily enters the aquatic food chain and is biomagnified as it accumulates in predatory fish such as swordfish, pike, and ocean tuna. Larger and more long-lived fish tend to contain more MeHg. Methylmercury contamination in fish can be significant; for example, the total mercury in the edible tissues of shark and swordfish can average as high as 1200 µg/kg (16). Marine mammal and fish consumption, while not the only sources of MeHg exposure, are extremely important routes of human exposure. This is particularly true for populations depending on fish as a primary food source.

Methylmercury is readily absorbed and distributed throughout the body, including the brain. In humans, MeHg brain levels are approximately six times higher than blood mercury levels (22). This is in contrast to rats, which have a brain-to-blood ratio of 0.06, and mice with a ratio of 1.20. These differences in brain MeHg accumulation have important implications for the extrapolation of human health guidelines from animal data. Methylmercury also readily crosses the placenta and appears to

accumulate in the fetus so that fetal mercury levels are greater than maternal blood mercury levels (23). The differential accumulation of MeHg in the brain and the fetus are important factors in defining developmental MeHg neurotoxicity.

Fetal Sensitivity to Methylmercury Exposure

Methylmercury is an excellent example of a known neurotoxicant that is also a prevalent and dangerous environmental pollutant. The effects of *in utero* exposure to MeHg are quite different from the effects associated with childhood or adult exposure (16). The fetus is more sensitive to the toxic effects of MeHg and severe effects have been found in the offspring of women showing little or no overt evidence of MeHg exposure (24–26). Catastrophic exposures in both Japan and Iraq provided evidence that mercury-exposed women delivered infants with severe behavioral and sensory deficits, including deafness and blindness, without expressing significant clinical signs or symptoms of mercury toxicity during pregnancy.

On both biological and neurobehavioral levels, there is strong evidence of fetal sensitivity to MeHg. Prenatal exposure to MeHg appears to result in a widespread pattern of adverse effects on brain development and organization (13,16,27–29). This generalized pattern of MeHg-induced injury to fetal brains is not seen in the adult brain, where MeHg exposure is characterized by localized lesions at specific neural sites (30). Postmortem studies from epidemic exposures in Japan and Iraq have revealed that MeHg significantly alters the normal migration of neurons to the cerebellar and cerebral cortices during brain development (27,28).

Examination of the brains of infants and animals exposed to MeHg *in utero* has revealed changes in neuronal migration and distribution patterns, cell loss (low neuronal abundance), and reduced brain size and gliosis (27,29,31,32). Hypotheses explaining MeHg-mediated developmental neurotoxicity include changes in intracellular cytoskeletal structure (33–35), oxidative stress (36–38), alterations to membrane function and signal transduction (39), decreased protein production (40), and changes in neurotransmission (41).

Neurobehavioral Effects in Human Infants

The disruptive effects of *in utero* MeHg exposure on brain development have been

associated with a broad range of neurobehavioral alterations in infancy and childhood. Published reports of studies involving mothers and infants come primarily from Japan, Iraq, Canada, and New Zealand. There are, however, additional reports of mother–infant pairs under study in the Seychelles Islands, Greenland, and the Faroe Islands (Table 1). Methylmercury exposure is usually estimated by blood or hair mercury levels, and it is generally accepted that the hair mercury concentration is 250 times that of the blood (16).

The catastrophic human exposures that occurred in Minamata and Niigata, Japan in the 1950s established that MeHg was fetotoxic (26,42). In the Minamata Bay study, 23 children believed to have been exposed to MeHg *in utero* showed evidence of mental retardation and cerebral palsy. During pregnancy, levels of Hg exposure were not monitored in the mothers of these children, but as a group these women showed little clinical evidence of MeHg toxicity. More subtle neurobehavioral deficits were not systematically studied in Japan; however, a general assessment of IQ in elementary and junior high-school students in the Minamata school district and a control district did not reveal any large differences in group performance (16). In the city of Minamata, a separate study found a significant correlation between level of MeHg in umbilical cord blood and occurrence of mental retardation (16).

In Iraq, an investigation of 29 mother–infant pairs was initiated after a serious outbreak of MeHg poisoning from consumption of contaminated bread (43). Results indicated a significant relationship between prenatal MeHg exposure and infant psychomotor retardation. Clear evidence of delays in attaining developmental milestones (e.g., motor and speech retardation) were evident in children with maternal hair MeHg levels less than 180 ppm. Neurological symptoms such as increased muscle tone and exaggerated deep tendon reflexes were primarily associated with maternal hair MeHg levels higher than 180 ppm. A subsequent report detailed the results of 84 mother–infant pairs (including the 29 discussed above), with peak maternal hair levels ranging from 0.4 to 640 ppm (44). Severe neurological symptoms (e.g., blindness, deafness, failure to walk, talk, or stand by over 4.5 years) were documented in five children. The lowest peak maternal hair level associated with severe neurological problems was 165 ppm (range 165–320 ppm). These reports were the first to docu-

Table 1. Human developmental effects of *in utero* exposure to MeHg.

Geographic location	Maternal MeHg exposure	Neurobehavioral effects	References
Japan	In Minamata, maternal samples taken 2 to 5 years after fetal delivery were elevated In Niigata, one mother had hair mercury levels of 293 ppm during pregnancy	Cerebral palsy, mental retardation, limb deformities, visual disorders, delayed speech, ataxia	Harada (26,42)
Iraq	Severe neurological deficits, with peak hair mercury concentrations from 165 to 320 ppm Mild deficits between 68 and 180 ppm	Mental and motor retardation, cerebral palsy, seizures, delayed speech, blindness, deafness	Marsh et al. (44)
Greenland	Mean blood value of 38.1 ppb	Reduced birth weight	Foldspang and Hansen (56)
Faroe Islands	Mean cord blood of 24.2 ppb Over 25% greater than 40 ppb	Neurobehavioral data not yet available	Grandjean et al. (57) Grandjean and Weihe (58)
New Zealand	Blood levels during pregnancy range from 6 to 86 ppb	Early sensorimotor deficits such as retarded walking, decreased scores on developmental tests	Kjellstrom et al. (53) Kjellstrom et al. (54)
Canada	Mean cord blood of 6 ppb; maternal hair 6 ppm	Abnormal deep tendon reflex in male infants	McKeown et al. (48)

ment more subtle impairments in children exposed to lower levels of MeHg during gestation and suggested a continuum of MeHg-related effects closely linked to maternal dose. Data from Iraq were reevaluated in an effort to determine a dose-response relationship between maternal hair levels of mercury and developmental effects (45). From this analysis, it was concluded that delayed onset of walking may occur at maternal hair levels of 10 to 20 ppm, which is equivalent to maternal blood mercury levels of 40 to 80 ppb.

The Iraq episode provided an opportunity for systematic follow-up of MeHg-exposed infants and children (46,47). Standard clinical and neurological tests including the Gesell Developmental Screening Exam were used as neurobehavioral assessment measures. Offspring effects ranged from hyperreflexia and delayed motor activity to microcephaly, cerebral palsy, and death. Follow-up studies at approximately 5 years of age indicated significant delays in psychomotor development and persistent pathological reflexes in a substantial number of children who did not display clinical signs during infancy. The clinical diagnoses of these children resembled minimal brain dysfunction (MBD) syndrome.

In Canada, a study of prenatal MeHg exposure in 234 Cree Indian infants and children did not find strong evidence of developmental abnormalities (48). Anthropometric measurements, neurological exams, and the Denver Developmental Scales were used as test measures with the children. The mean maternal hair level in this study was 6 ppm. No effects on physical development were noted for either males or females. Significant neurobehav-

ioral effects were limited to the finding that maternal exposure to MeHg was related to abnormal muscle tone (deep tendon reflex) in male infants. No effects of exposure to MeHg were noted in female infants and the authors note the questionable clinical significance of this sex-specific finding. Animal data, however, would suggest that there may be sex-related effects of prenatal MeHg exposure, with males generally showing a greater sensitivity to *in utero* exposure (29,49-52).

Studies of children prenatally exposed to MeHg through maternal fish consumption have been conducted in New Zealand (53). Approximately 1000 women were identified as frequent fish consumers and of these, 73 were identified as having maternal hair concentrations above 6 ppm (range 6-86 ppm). At 4 years of age, 31 offspring from this group were assessed with the Denver Developmental Screening Test. Evidence of a significant increase in the risk of early sensorimotor dysfunction was documented in the MeHg-exposed group. A dose-response relationship was established between mean maternal hair MeHg levels and performance on the Denver Developmental Screening Test.

A subsequent study evaluated 61 of the original 73 high-dose children, at 6 to 7 years of age, on multiple assessments that included tests of intelligence (WISC-R) and language development (TOLD) (54). Results showed that exposed children who scored poorly on the Denver Developmental Screening Test at 4 years of age tended to have decreased scores on the WISC-R intelligence test later in childhood. These neurobehavioral effects were associated with maternal blood MeHg levels of only 20 to 80 ppb.

Elevated levels of blood mercury in women of childbearing age have been found in polar Inuit natives in Northern Greenland (55). The Inuits' reliance on whale meat as a dietary mainstay is believed to be the primary source of exposure. Of the women tested, 84% had blood MeHg levels that exceeded the provisional limit set by the WHO (23 ppb). As expected, the fetal levels of MeHg in cord blood (average value of 80.2 ppb) were higher than maternal blood levels (average value of 38.1 ppb). An examination of the relationship between cord blood MeHg and birth weight revealed that decreased birth weights were associated with higher levels of fetal MeHg exposure (56). Although information on the neurobehavioral status of these children is not currently available, the cord blood MeHg levels are in the range of values that have been associated with psychomotor retardation (53).

Samples of cord blood and maternal hair also were collected and assayed for mercury from women living in the Faroe Islands (57,58). With a sample size of 1000 infants, the mean mercury concentration in cord blood was 24.2 ppb, and over 25% of the samples were above 40 ppb. This clearly exceeds the WHO provisional limit for potential health effects and confirms elevated *in utero* MeHg exposure. Faroese children as well as the Inuit children of Northern Greenland should be considered at risk for neurobehavioral alterations associated with *in utero* MeHg exposure.

In summary, a review of the human data on the developmental effects of MeHg exposure indicates that maternal hair levels of 10 to 20 ppm are potentially harmful to

fetal development. This is equivalent to maternal blood levels of 40 to 80 ppb of mercury, assuming a mercury hair-to-blood ratio of 250. To determine a LOAEL, the blood mercury levels must be converted to an estimate of daily consumption (i.e., dose) that would result in the equivalent blood levels. Based on kinetic modeling of MeHg, it is estimated that the long-term daily consumption of 1 µg of mercury will result in blood mercury levels of 1 µg/liter or 1 ppb (16). Thus, the consumption of 40 µg of mercury per day would result in a blood mercury level of 40 ppb; which for a 62-kg woman would be equivalent to 0.645 µg/kg/day. If the typically used uncertainty factor of 10 is used to account for sensitive individuals (in this case fetal development), the resulting NOAEL or RfD would be 0.06 µg/kg/day.

Neurobehavioral Effects in Animals

As discussed above, human exposure to MeHg during prenatal development results in a continuum of effects ranging from blindness, deafness, seizures, abnormal reflexes, and retarded motor development, to far more subtle learning, memory, and psychological effects (5,9,25,26,47,59). Adverse effects of MeHg exposure can occur at human brain levels estimated to be as low as 0.3 ppm (45).

While animal studies using high levels of MeHg exposure have produced effects similar to those of humans, there have been few efforts to characterize the more subtle effects of low-dose exposure to MeHg during development. Early effects of *in utero* exposure have been documented in nonhuman primate infants. As part of a larger study examining the maternal reproductive and offspring developmental effects of chronic exposure to MeHg, female *Macaca fascicularis* monkeys were exposed to daily doses of MeHg throughout pregnancy (0, 50, 70, or 90 µg/kg/day). Maternal blood MeHg levels averaged 1.28, 1.62, and 2.03 ppm, respectively, for the three treated groups. Maternal blood MeHg levels above 1.5 ppm were associated with a significant decrease in number of viable births (60).

During infancy, effects of prenatal exposure were found on measures of cognitive and social development in the offspring of the MeHg-exposed monkeys. *In utero* exposure to MeHg was related to delayed attainment of object permanence (61), deficits in visual recognition memory (62,63), and abnormal social behavior

(64). These results, frequently based on test procedures developed for use with human infants, show that *in utero* exposure to MeHg is related to delays in the attainment of important cognitive milestones. The social behavior of the MeHg-exposed infants in established play groups was characterized by a significant decrease in play behavior and an increase in nonsocial behavior. Exposed infants were less likely to engage in species-appropriate play behavior and spent more time alone, distancing themselves from other monkeys in the group. In this same group of animals, a latent effect of prenatal exposure to MeHg indicated pubertal growth retardation in exposed males (49). Exposed males exhibited significantly decreased weight gain during the juvenile stages of growth (3–5 years of age) but did catch up to the average weight of control males by early adulthood. Subsequent studies in adulthood with these animals have found very slight effects on an intermittent schedule of reinforcement (fixed-interval/fixed-ratio) (65). In this group of animals, overall study results do not support long-term deficits in adult learning and memory abilities (66). However, preliminary results indicate there may be deficits in adult visual function. Results from this group of monkeys indicate that developmental effects are seen at maternal exposures of 50 µg/kg MeHg/day.

Confirmation of the developmental effects of MeHg exposure is evident from Canadian studies in which monkeys were exposed to MeHg either post- or pre- and postnatally (birth to 7 years of age) at 25 or 50 µg/kg/day. In one study, infants were tested on a fixed-interval operant learning task. Exposed infants showed slight alterations in performance, indicating a possible disruption of time perception (67). In the same study, MeHg exposure was not related to learning impairments on discrimination reversal tests. Developmental exposure to MeHg was also shown adversely to affect visual, auditory, and somatosensory function in monkeys (67–71). Visual psychophysical studies with these monkeys have shown treatment-related deficits in spatial contrast sensitivity and facilitation of low-luminance temporal contrast sensitivity (69). Subsequent studies have described overt sensory-motor deficits (i.e., lack of coordination in exercise cages) and a loss of vibration sensitivity (70–72). These effects were observed at the lowest doses tested

(25 µg/kg), which is characteristic of many studies with MeHg.

In rodents, one of the most frequent findings related to prenatal MeHg exposure is an increased rate of intrauterine death (10). Studies on the developmental effects of MeHg on rats (Table 2) and mice (Table 3) have provided a means of examining specific hypotheses regarding the mechanisms of action of MeHg. These studies also provide information on the levels of maternal exposure at which no adverse effects are observed in the offspring. Typically, pregnant rats or mice were dosed with MeHg for a restricted period of time during gestation. To compensate for the short exposure periods, relatively high doses of MeHg were often used. The most common neurological deficit observed was altered locomotion or exploratory behavior (73–78). These findings are consistent with results from high-exposure human studies which revealed significant delays in aspects of motor development such as crawling, standing, and walking. Learning deficits have also been observed following developmental exposure to MeHg (77–81).

The most comprehensive study to assess the developmental effects of MeHg in rats was done as part of the Collaborative Behavioral Teratology Study (78). In this multilaboratory study, using the same testing protocol, pregnant rats were exposed to either 2 or 6 mg/kg of MeHg on days 6 through 9 of gestation. These studies found dose-related changes in behavior characterized by increased levels of activity and impaired learning of auditory startle habituation in exposed pups. Lower doses of MeHg were not examined, so a NOAEL could not be determined. Two studies have assessed the effects of low-level prenatal exposure to MeHg using schedule-controlled operant behavior (82,83). In these studies, rats were dosed with either 0.005, 0.01, 0.05, or 2.0 mg/kg of MeHg during days 6 through 9 of gestation. The offspring were then tested on a differential reinforcement of high rate schedule. This task requires the subject to respond to a lever a specified number of times within a fixed period to receive a reinforcement (e.g., two responses are required within 1 sec). No adverse effects were observed at a dose of 0.005 mg/kg. All other treated groups had reduced success rates. Although no blood or brain mercury levels were reported, it is estimated that the brain Hg levels were as

Table 2. Effects of *in utero* MeHg exposure in rats.

Dose, mg/kg	Exposure	Noted effects	NOAEL	Reference
0.005, 0.01, or 0.05	GD 6–9	Differences in DRH operant testing noted in 0.01 and 0.05 dose groups.	0.005 mg/kg dose group	Bornhausen et al. (83)
0, .25, 1.25, 2.50, or 5	GD 6–15	No live offspring in 5 mg/kg dose group; 2.5 mg/kg group displayed impairment in all preweaning measures, locomotion, open field and startle response performance; 1.25 mg/kg group showed deficits only in swimming	0.25 mg/kg dose group	Geyer et al. (73)
0, .05, 0.5, or 5	GD 0, 7, or 14	No noted differences in litter size, birth weight, gross appearance, or operant behavior	5 mg/kg dose group	Hughes and Sparber (101)
0, .05, or 2.0	GD 6–9	Showed dose-dependent decreases in learning using the DRH operant test. No noted differences in general motility or motor coordination	No NOAEL	Musch et al. (82)
0, 2 or 6	GD 6–9	Auditory startle habituation increased, increased activity measures, alteration in visual discrimination	No NOAEL	Buelke-Sam et al. (78)
0 or 8	GD 8	Stereotyped sniffing elicited in treated only; no changes in locomotor activity, altered passive avoidance response	No NOAEL	Cuomo et al. (80)
0, 5, or 8	GD 8 or 15	Decreased maternal weight gain at high dose, decreased neonatal activity, reduced acquisition two-way avoidance	No NOAEL	Eccles and Annau (81,102)
0, 2, or 6	GD 6–9	Both dose groups showed reduced postweaning figure 8 activity, increased Biel water maze time, errors and proportion of trial failures in addition to delayed developmental growth	No NOAEL	Vorhees (77)
0, 10 (IP)	GD 18	Altered brain cellular arrangement and neuronal migration	No NOAEL	Geelen et al. (32)
0 or 5	GD 7	Altered visual-evoked potentials	No NOAEL	Dyer et al. (103)
0 or 2.5	Throughout	Altered visual-evoked potentials	No NOAEL	Zenick (104)
0 or 2.5	Throughout	Increased errors in water T-maze; no motor impairment	No NOAEL	Zenick (105)

GD, gestational day(s).

Table 3. Effects of *in utero* MeHg exposure in mice.

Dose, mg/kg	Exposure	Noted effects	NOAEL	Reference
0, 1, 2, 3, 5, or 10	GD 8	Effects noted in animals dosed 3.0 mg/kg or higher in two-way shuttle box, active and passive avoidance, litter size, no motor impairment	2 mg/kg dose group	Hughes and Annau (106)
20, 25, or 30	GD 14, 15, or 16	Offspring at 25 and 30 mg/kg did not survive; 20 mg/kg group showed neurologic disturbances, righting problems	No NOAEL	Inouye et al. (74)
0, 6, 8, or 12	GD 10	All dose groups showed decreased exploratory behavior, rearing, urination, and increased backing	No NOAEL	Su and Okita (75)
0, 2, 4, or 8	GD 6–13	Retarded growth, embryolethal and teratogenic in 129 (when dosed days 9–13), teratogenic in A/J (dosed days 9–13)	No NOAEL	Spyker and Smithberg (107)
0 or 8	GD 7 or 9	Decreased exploratory behavior, increased backing; demonstrated neuromuscular impairment while swimming	No NOAEL	Spyker et al. (76)

GD, gestational day(s).

low as 0.04 ppm, well below levels thought to produce adverse effects in humans.

In summary, results from monkey studies indicate that a LOAEL for developmental effects is evident at a dose of 25 µg/kg/day of MeHg. Rodent studies indicate effects may be observed at doses as low as 10 µg/kg/day of MeHg, with effects replicated at 50 µg/kg/day. Using the results from the monkey studies, a NOAEL of 2.5 µg/kg/day is derived using the standard procedure of dividing the LOAEL by a factor of 10. To convert a NOAEL derived from animal studies to one for humans, an uncertainty factor of 10 is typically used to account for interspecies differences. An additional uncertainty factor of 10 is then applied to account for sensitive individuals in the human population. Using a factor of 10 is appropriate given the sensitivity of the developing nervous system to

the adverse effects of MeHg. The above risk analysis indicates that a more conservative RfD for MeHg exposure would be 0.025 µg/kg/day, which would provide a level of safety for the developing nervous system.

Neurobehavioral Effects of Other Neurotoxic Compounds

Methylmercury clearly is not the only compound that can adversely affect the developing nervous system. Lead is one of the best studied environmental neurotoxicants and is a good example of the problems associated with understanding the effects of very low levels of exposure (84–86). The widespread exposure of children to lead made it possible to perform numerous human epidemiology studies that convincingly demonstrated that low-level exposure is harmful to the developing nervous system (87). Animal studies, particularly those in

monkeys, confirmed that low-level exposure to lead had adverse developmental effects (88). The consequences of the deleterious effects of lead on normal childhood development are just beginning to be examined (85,89).

Another important widespread environmental contaminant and neurotoxicant is the lipid-soluble polychlorinated biphenyls (PCBs). This family of over 200 chemicals was used primarily as insulators in electrical equipment. Production of PCBs was banned in the 1970s following recognition of their toxicity and environmental persistence. These lipid-soluble compounds are mobilized during pregnancy, thus exposing the infant *in utero* and are also readily excreted in breast milk during lactation. The neurotoxic effects of *in utero* exposure to PCBs are well documented and have been carefully reviewed (90–94). It is

interesting to note that the effects of PCBs appeared to be more related to *in utero* exposure than to postnatal exposure through breast milk.

The voluntary consumption of neurotoxic substances during pregnancy can also result in an array of neurobehavioral effects. Alcohol consumption during pregnancy produces a well-documented syndrome of adverse effects on the nervous system that range from subtle deficits in learning and memory to severe developmental disorders (95–97). Animal studies with alcohol have made important contributions to the understanding of fetal alcohol effects and demonstrate the comparability of human and animal findings (98). Studies on the long-term impact of prenatal alcohol exposure demonstrate the individual and societal consequences of early neurotoxicant exposure (95,97,99). Other drugs such as cocaine also affect nervous system development (100).

Conclusions and Recommendations

Methylmercury is a compound worthy of scientific and societal concern. It is clear that MeHg is a widespread environmental contaminant and a potent neurotoxicant that adversely affects the developing nervous system. Mercury continues to be released into the environment by both nat-

ural and human-generated sources. It is readily converted to MeHg and accumulates in the food supply, primarily in fish and marine mammals. MeHg is readily absorbed and distributed throughout the body, including the brain and the fetus. Fetal exposure appears to be at a level that is greater than maternal blood levels. Studies of humans exposed to elevated levels of MeHg clearly demonstrate its neurotoxic potential. Animal studies using rodents and nonhuman primates have confirmed the neurotoxic potential of MeHg. However, research into cellular and molecular mechanisms has yet to produce an understanding of MeHg sufficient to allow accurate prediction of its neurotoxicity. Furthermore, human and animal studies on the neurobehavioral effects of developmental MeHg exposure have not determined a level of exposure that is convincingly harmless to the developing fetus.

In many ways, our understanding of the neurotoxic potential of MeHg is similar to that of lead 20 years ago; MeHg is a known neurotoxicant at high levels of exposure but there is little understanding of its effects at lower levels of exposure. The failure to adequately characterize the functional effects of low-level MeHg exposure has compromised the formulation of a sound policy regarding the safe levels of MeHg exposure, particularly for pregnant women or women of child-bearing age.

Examination of the results of human studies on the effects of MeHg indicate that maternal hair levels of 10 to 20 ppm may result in adverse effects on fetal outcome. Making the appropriate assumptions and calculations, a level of exposure not expected to be hazardous (RfD) would be 0.06 µg/kg/day. Evaluation of results from animal studies on the developmental effects of MeHg provided an estimated RfD of 0.025 µg/kg/day. The human and animal RfDs are in very good agreement.

Given the current state of knowledge with regard to MeHg exposure, the following recommendations are offered:

- reduce environmental release of all forms of mercury;
- consider restricting the global production and sale of mercury;
- strongly advise pregnant women and women of child bearing age to limit their exposure to sources of MeHg;
- establish an RfD (reference dose) for MeHg of 0.025 to 0.06 µg/kg/day;
- continue research to determine a level of MeHg exposure that would not harm the developing nervous system;
- continue research to understand the underlying molecular mechanisms of action of MeHg;
- assess the long-term neurodegenerative effects of developmental MeHg exposure.

REFERENCES

1. OTA. Neurotoxicity: Identifying and Controlling Poisons of the Nervous System. Washington:U.S. Congress, Office of Technology Assessment, 1990.
2. NRC. Environmental Neurotoxicology. National Research Council, 1991.
3. Landrigan PJ, Graham DG, Thomas RD. Strategies for the prevention of environmental neurotoxic illness. *Environ Res* 61:157–163 (1993).
4. WHO. Inorganic Mercury Environmental Health Criteria 118. Geneva:World Health Organization, 1991.
5. Burbacher TM, Rodier PM, Weiss B. Methylmercury developmental neurotoxicity: a comparison of effects in humans and animals. *Neurotoxicol Teratol* 12:191–202 (1990).
6. Chang L, Reuhl K. Mercury in human and animal health. In: *Trace Elements in Health: A Review of Current Issues* (Rose J, ed). Boston:Butterworths, 1983;132–149.
7. Chang LW. Neurotoxic effects of methylmercury intoxication—a review. *Environ Res* 14:329–373 (1987).
8. Clarkson TW, Hursh JB, Sager PR, Syversen TLM. Mercury. In: *Biological Monitoring of Toxic Metals* (Clarkson TW, Friberg L, Nordberg GF, Sager PR, eds). New York:Johns Hopkins Press, 1988;199–246.
9. Clarkson TW. Mercury: major issues in environmental health. *Environ Health Perspect* 100:31–38 (1992).
10. Eccles CV, Annau Z. *The Toxicity of Methylmercury*. Baltimore:Johns Hopkins University Press, 1987.
11. ATSDR. Toxicological Profile for Mercury. Atlanta:Agency for Toxic Substances and Disease Registry, 1994.
12. Fitzgerald WF, Clarkson TW. Mercury and monomethylmercury: present and future concerns. *Environ Health Perspect* 96:159–166 (1991).
13. Reuhl KR, Chang LW. Effects of methylmercury on the development of the nervous system: a review. *Neurotoxicology* 1:21–55 (1979).
14. Stern AH. Re-evaluation of the reference dose for methylmercury and assessment of current exposure levels. *Risk Anal* 13:355–364 (1993).
15. U.S. EPA. U Methylmercury—IRIS (Integrated Risk Information System). Washington:U.S. Environmental Protection Agency, 1992.
16. WHO. Methylmercury. Environmental Health Criteria 101. Geneva:World Health Organization, 1990.
17. WHO. Mercury—Environmental Aspects. Environmental Health Criteria 86. Geneva:World Health Organization, 1989.
18. Pfeiffer WC, de-Lacerda LD, Malm O, Souza CM, da-Silveira EG, Bastos WR. Mercury concentrations in inland waters of gold-mining areas in Rondonia, Brazil. *Sci Total Environ* 88:233–240 (1989).
19. Brosset C. The mercury cycle. *Water Air Soil Poll* 16:253–255 (1981).
20. Pfeiffer WC, Drude de Lacerda L. Mercury inputs into the Amazon Region, Brazil. *Environ Technol Lett* 9:325–330 (1988).
21. Drude de Lacerda L, Pfeiffer WC, Ott AT, Gloria da Silveira E. Mercury contamination in the Madeira River, Amazon—Hg inputs to the environment. *Biotropica* 21:91–93 (1989).
22. Magos L. The absorption, distribution and excretion of methyl

- mercury. In: The Toxicity of Methylmercury (Eccles CV, Annau Z, eds). Baltimore:Johns Hopkins University Press, 1987;24-44.
23. Suzuki T, Yonemoto J, Satoh H, Naganuma A, Imura N, Kigawa T. Normal organic and inorganic mercury levels in the human fetoplacental system. *J Appl Toxicol* 4:249-252 (1984).
 24. Marsh DO. Dose-response relationships in humans: methyl mercury epidemics in Japan and Iraq. In: The Toxicity of Methyl Mercury (Eccles CV, Annau Z, eds). Baltimore:Johns Hopkins University Press, 1987;45-53.
 25. Marsh DO, Clarkson TW, Cox C, Myers GJ, Amin ZL, Al TS. Fetal methylmercury poisoning. Relationship between concentration in single strands of maternal hair and child effects. *Arch Neurol* 44:1017-1022 (1987).
 26. Harada Y. Congenital Minamata disease. In: Minamata Disease: Methyl Mercury Poisoning in Minamata and Niigata (Tsubak R, Irukayama K, eds). Tokyo:Kodansha, 1977;209-239.
 27. Choi BH, Lapham LW, Amin-Zaki L, Saleem T. Abnormal neuronal migration, deranged cerebral cortical organization, and diffuse white matter astrocytosis of fetal human brain: a major effect of methylmercury poisoning *in utero*. *J Neuropathol Exp Neurol* 37:719-733 (1978).
 28. Choi BH. Methylmercury poisoning of the developing nervous system: I. Pattern of neuronal migration in the cerebral cortex. *Neurotoxicology* 7:591-600 (1986).
 29. Matsumoto H, Koya G, Takeuchi T. Fetal Minamata disease. *J Neuropathol Exer Neurol* 24:563-574 (1965).
 30. Hamada R, Yoshida Y, Nomoto M, Osame M, Igata A, Mishima I, Kuwano A. Computed tomography in fetal methylmercury poisoning. *J Toxicol Clin Toxicol* 31:101-106 (1993).
 31. Mottet NK, Shaw CM, Burbacher TM. Health risks from increases in methylmercury exposure. *Environ Health Perspect* 63:133-140 (1985).
 32. Geelen JA, Dormans JA, Verhoef A. The early effects of methylmercury on the developing rat brain. *Acta Neuropathol Berl* 80:432-438 (1990).
 33. Sager PR. Selectivity of methylmercury effects on cytoskeleton and mitotic progression in cultured cells. *Toxicol Appl Pharmacol* 94:473-486 (1988).
 34. Sager PR, Matheson DW. Mechanisms of neurotoxicity related to selective disruption of microtubules and intermediate filaments. *Toxicology* 49:479-492 (1988).
 35. Vogel DG, Margolis RL, Mottet NK. Analysis of methylmercury binding sites on tubulin subunits and microtubules. *Pharm Toxicol* 64:196-201 (1989).
 36. Bondy SC, McKee M. Prevention of chemically induced synaptosomal changes. *J Neurosci Res* 25:229-235 (1990).
 37. LeBel CP, Ali SF, McKee M, Bondy SC. Organometal-induced increases in oxygen reactive species: the potential of 2',7'-dichlorofluorescein diacetate as an index of neurotoxic damage. *Toxicol Appl Pharmacol* 104:17-24 (1990).
 38. Sarafian T, Verity MA. Oxidative mechanisms underlying methylmercury neurotoxicity. *Int J Dev Neurosci* 9:147-153 (1991).
 39. Bartolome JV, Kavlock RJ, Cowdery T, Orband-Miller L, Slotkin TA. Development of adrenergic receptor binding sites in brain regions of the neonatal rat: effects of prenatal or postnatal exposure to methylmercury. *Neurotoxicology* 8:1-14 (1987).
 40. Syversen TL, Totland G, Flood PR. Early morphological changes in rat cerebellum caused by a single dose of methylmercury. *Arch Toxicol* 47:101-111 (1981).
 41. Slotkin TA, Bartolome J. Biochemical mechanisms of developmental neurotoxicity of methylmercury. *Neurotoxicology* 8:65-84 (1987).
 42. Harada M. Congenital Minamata disease: intrauterine methylmercury poisoning. *Teratology* 18:285-288 (1978).
 43. Marsh DO, Myers GJ, Clarkson TW, Amin-Zaki L, Tikriti S, Majeed MA. Fetal methylmercury poisoning: clinical and toxicological data on 29 cases. *Ann Neurol* 7:348-353 (1980).
 44. Marsh DO, Myers GJ, Clarkson TW, Amin-Zaki L, Tikriti S, Majeed MA, Dabbagh AR. Dose-response relationship for human fetal exposure to methylmercury. *Clin Toxicol* 18:1311-1318 (1981).
 45. Cox C, Clarkson TW, Marsh DO, Amin ZL, Tikriti S, Myers GG. Dose-response analysis of infants prenatally exposed to methyl mercury: an application of a single compartment model to single-strand hair analysis. *Environ Res* 49:318-332 (1989).
 46. Amin-Zaki L, Majeed MA, Elhassani SB, Clarkson TW, Greenwood MR, Doherty RA. Prenatal methylmercury poisoning. Clinical observations over five years. *Am J Dis Child* 133:172-177 (1979).
 47. Amin-Zaki L, Majeed MA, Greenwood MR, Elhassani SB, Clarkson TW, Doherty RA. Methylmercury poisoning in the Iraqi suckling infant: a longitudinal study over five years. *J Appl Toxicol* 1:210-214 (1981).
 48. McKeown EGE, Ruedy J, Neims A. Methyl mercury exposure in northern Quebec. II. Neurologic findings in children. *Am J Epidemiol* 118:470-479 (1983).
 49. Grant-Webster KS, Burbacher TM, Mottet NK. Puberal growth retardation in primates: a latent effect of *in utero* exposure to methylmercury. *Toxicologist* 12:310 (1992).
 50. Sager PR, Aschner M, Rodier PM. Persistent, differential alterations in developing cerebellar cortex of male and female mice after methylmercury exposure. *Brain Res* 314:1-11 (1984).
 51. Hirayama K, Yasutake A. Sex and age differences in mercury distribution and excretion in methylmercury-administered mice. *J Toxicol Environ Health* 18:49-60 (1986).
 52. Hirayama K, Yasutake A, Inoue M. Effect of sex hormones on the fate of methylmercury and on glutathione metabolism in mice. *Biochem Pharmacol* 36:1919-1924 (1987).
 53. Kjellstrom T, Kennedy P, Wallis S, Mantell C. Physical and mental development of children with prenatal exposure to mercury from fish. Stage 1: Preliminary tests at age 4. Sweden: National Swedish Environmental Protection Board, 1986.
 54. Kjellstrom T, Kennedy P, Wallis S, Stewart A, Friberg L, Lind B, Wutherspoon P, Mantell C. Physical and mental development of children with prenatal exposure to mercury from fish. Stage 2: interviews and psychological tests at age 6. Sweden: National Swedish Environmental Protection Board, 1989.
 55. Hansen JC, Tarp U, Bohm J. Prenatal exposure to methyl mercury among Greenlandic polar Inuits. *Arch Environ Health* 45:355-358 (1990).
 56. Foldspang A, Hansen JC. Dietary intake of methylmercury as a correlate of gestational length and birth weight among newborns in Greenland. *Am J Epidemiol* 132:310-317 (1990).
 57. Grandjean P, Weihe P, Jorgensen PJ, Clarkson T, Cernichiari E, Videro T. Impact of maternal seafood diet on fetal exposure to mercury, selenium, and lead. *Arch Environ Health* 47:185-195 (1992).
 58. Grandjean P, Weihe P. Neurobehavioral effects of intrauterine mercury exposure: potential sources of bias. *Environ Res* 61:176-183 (1993).
 59. Amin-Zaki L, Elhassani S, Majeed MA, Clarkson TW. Perinatal methylmercury poisoning in Iraq. *Am J Dis Child* 130:1070-1076 (1976).
 60. Burbacher TM, Mohamed MK, Mottet NK. Methylmercury effects on reproduction and offspring size at birth. *Reprod Toxicol* 1:267-278 (1988).
 61. Burbacher TM, Grant KS, Mottet NK. Retarded object permanence development in methylmercury exposed *Macaca fascicularis* infants. *Devel Psychol* 22:771-776 (1986).
 62. Gunderson VM, Grant KS, Burbacher TM, Fagan JF III, Mottet NK. The effect of low-level prenatal methylmercury exposure on visual recognition memory in infant crab-eating macaques. *Child Dev* 57:1076-1083 (1986).
 63. Gunderson VM, Grant WKS, Burbacher TM, Mottet NK. Visual recognition memory deficits in methylmercury-exposed *Macaca fascicularis* infants. *Neurotoxicol Teratol* 10:373-379 (1988).
 64. Burbacher TM, Sackett GP, Mottet NK. Methylmercury effects on the social behavior of *Macaca fascicularis* infants.

- Neurotoxicol Teratol 12:65–71 (1990).
65. Gilbert SG, Rice DC, Burbacher TM. Fixed interval-fixed ratio performance in adult monkeys exposed *in utero* to methylmercury. Neurotoxicol Teratol (in press).
66. Gilbert SG, Burbacher TM, Rice DC. Effects of *in utero* methylmercury exposure on a spatial delayed alternation task in monkeys. Toxicol Appl Pharmacol 123:130–136 (1993).
67. Rice DC. Effects of pre- plus postnatal exposure to methylmercury in the monkey on fixed interval and discrimination reversal performance. Neurotoxicology 13:443–452 (1992).
68. Rice DC, Gilbert SG. Early chronic low-level methylmercury poisoning in monkeys impairs spatial vision. Science 216:759–761 (1982).
69. Rice DC, Gilbert SG. Effects of developmental exposure to methylmercury on spatial and temporal visual function in monkeys. Toxicol Appl Pharmacol 102:151–163 (1990).
70. Rice DC, Gilbert SG. Delayed somatosensory deficits in monkeys exposed developmentally to methylmercury. International Neurotoxicology Association, 1993.
71. Rice DC, Gilbert SG. Effects of developmental methylmercury exposure or life time lead exposure on vibration sensitivity function in monkeys. Toxicol Appl Pharmacol (in press).
72. Rice DC. Delayed neurotoxicity in monkeys exposed developmentally to methylmercury. Neurotoxicology 10:645–650 (1989).
73. Geyer MA, Butcher RE, Fite K. A study of startle and locomotor activity in rats exposed prenatally to methylmercury. Neurobehav Toxicol Teratol 7:759–765 (1985).
74. Inouye M, Murao K, Kajiwara Y. Behavioral and neuropathological effects of prenatal methylmercury exposure in mice. Neurobehav Toxicol Teratol 7:227–232 (1985).
75. Su MQ, Okita GT. Behavioral effects on the progeny of mice treated with methylmercury. Toxicol Appl Pharmacol 38:195–205 (1976).
76. Spyker JM, Sparber SB, Goldberg AM. Subtle consequences of methylmercury exposure: behavioral deviations in offspring of treated mothers. Science 177:621–623 (1972).
77. Vorhees C. Behavioral effects of prenatal methylmercury in rats: a parallel trial to the collaborative behavioral teratology study. Neurobehav Toxicol Teratol 7:717–725 (1985).
78. Buelke-Sam J, Kimmel CA, Adams J, Nelson CJ, Vorhees CV, Wright DC, St OV, Korol BA, Butcher RE, Geyer MA. Collaborative behavioral teratology study: results. Neurobehav Toxicol Teratol 7:591–624 (1985).
79. Schalock RL, Brown WJ, Kark RA, Menon NK. Perinatal methylmercury intoxication: behavioral effects in rats. Dev Psychobiol 14:213–219 (1981).
80. Cuomo V, Ambrosi L, Annau Z, Cagiano R, Brunello N, Racagni G. Behavioural and neurochemical changes in offspring of rats exposed to methylmercury during gestation. Neurobehav Toxicol Teratol 6:249–254 (1984).
81. Eccles CU, Annau Z. Prenatal methylmercury exposure: II. Alterations in learning and psychotropic drug sensitivity in adult offspring. Neurobehav Toxicol Teratol 4:377–382 (1982).
82. Musch HR, Bornhausen M, Kreigel H, Greim H. Methylmercury chloride induces learning deficits in prenatally treated rats. Arch Toxicol 40:103–108 (1978).
83. Bornhausen M, Musch HR, Greim H. Operant behavior performance changes in rats after prenatal methylmercury exposure. Toxicol Appl Pharmacol 56:305–310 (1980).
84. Needleman HL, Bellinger D. The health effects of low level exposure to lead. Annu Rev Public Health 12:111–140 (1991).
85. Needleman HL. The future challenge of lead toxicity. Environ Health Perspect 89:85–89 (1990).
86. Needleman HL. What can the study of lead teach us about other toxicants? Environ Health Perspect 86:183–189 (1990).
87. Needleman HL, Gatsonis CA. Low-level lead exposure and the IQ of children. A meta-analysis of modern studies. JAMA 263:673–678 (1990).
88. Rice DC. Behavioral impairment produced by developmental lead exposure: evidence from primate research. In: Human Lead Exposure (Needleman HL, ed). Boca Raton, FL: CRC Press, 1992:137–154.
89. Needleman HL. The persistent threat of lead: medical and sociological issues. Curr Probl Pediatr 18:697–744 (1988).
90. Rogan WJ, Gladen BC. Neurotoxicology of PCBs and related compounds. Neurotoxicology 13:27–35 (1992).
91. Tilson HA, Jacobson JL, Rogan WJ. Polychlorinated biphenyls and the developing nervous system: cross-species comparisons. Neurotoxicol Teratol 12:239–248 (1990).
92. Rogan WJ, Gladen BC, McKinney JD, Carreras N, Hardy P, Thullen J, Tinglestad J, Tully M. Neonatal effects of transplacental exposure to PCBs and DDE. J Pediatr 109:335–341 (1986).
93. Jacobson JL, Jacobson SW, Humphrey HE. Effects of *in utero* exposure to polychlorinated biphenyls and related contaminants on cognitive functioning in young children. J Pediatr 116:38–45 (1990).
94. Jacobson JL, Jacobson SW, Humphrey HE. Effects of exposure to PCBs and related compounds on growth and activity in children. Neurotoxicol Teratol 12:319–326 (1990).
95. Coles CD. Impact of prenatal alcohol exposure on the newborn and the child. Clin Obstet Gynecol 36:255–266 (1993).
96. Jacobson JL, Jacobson SW, Sokol RJ, Martier SS, Ager JW, Kaplan-Estrin MG. Teratogenic effects of alcohol on infant development. Alcohol Clin Exp Res 17:174–183 (1993).
97. Streissguth AP. Fetal alcohol syndrome: early and long-term consequences. Nida Res Monogr 119:126–130 (1992).
98. Driscoll CD, Streissguth AP, Riley EP. Prenatal alcohol exposure: comparability of effects in humans and animal models. Neurotoxicol Teratol 12:231–237 (1990).
99. Streissguth AP. Prenatal alcohol-induced brain damage and long-term postnatal consequences: introduction to the symposium. Alcohol Clin Exp Res 14:648–649 (1990).
100. Ellis JE, Byrd LD, Sexson WR, Patterson-Barnett CA. *In utero* exposure to cocaine: a review. South Med J 86:725–731 (1993).
101. Hughes JA, Sparber SB. *d*-Amphetamine unmasks postnatal consequences of exposure to methylmercury *in utero*: methods for studying behavioral teratogenesis. Pharmacol Biochem Behav 8:365–375 (1978).
102. Eccles CU, Annau Z. Prenatal methyl mercury exposure: I. Alterations in neonatal activity. Neurobehav Toxicol Teratol 4:371–376 (1982).
103. Dyer RS, Eccles CU, Annau Z. Evoked potential alterations following prenatal methyl mercury exposure. Pharmacol Biochem Behav 8:137–141 (1978).
104. Zenick H. Evoked potential alterations in methylmercury chloride toxicity. Pharmacol Biochem Behav 5:253–255 (1976).
105. Zenick H. Behavioral and biochemical consequences in methylmercury chloride toxicity. Pharmacol Biochem Behav 2:709–713 (1974).
106. Hughes JA, Annau Z. Postnatal behavioral effects in mice after prenatal exposure to methylmercury. Pharmacol Biochem Behav 4:385–391 (1976).
107. Spyker JM, Smithberg M. Effects of methylmercury on prenatal development in mice. Teratology 5:181–190 (1972).